http://dx.doi.org/10.7124/bc.000AC5

## Epigenetic, transcriptional and splicing changes in the glioblastoma marker genes *CHI3L1* and *MGMT* during the acquisition of temozolomide resistance

O.V. Anopriienko<sup>1,2</sup>, M.K. Shuvalova<sup>2</sup>, P.O. Areshkov<sup>1</sup>, A.R. Shloma<sup>1</sup>, K.I. Solomiana<sup>1</sup>, I.Ya. Skrypkina<sup>1,2</sup>

<sup>1</sup> Institute of Molecular Biology and Genetics, NAS of Ukraine 150, Akademika Zabolotnoho Str., Kyiv, Ukraine, 03143

<sup>2</sup> The State Research Institution «Kyiv Academic University»

36, Akademika Vernadsky Blvd., Kyiv, Ukraine, 03142

o.v.anoprienko@imbg.org.ua

Background/Aim. CHI3L1 (chitinase 3-like protein 1) is a secretory glycoprotein highly upregulated in glioblastoma (GBM) and is recognized as a molecular marker of tumor mesenchymal subtype. However the gene's short transcriptional isoform (CHI3L1A8) is poorly studied regarding its biological role in GBM. O6-methylguanine-DNA methyltransferase (MGMT) promoter methylation is considered a prognostic marker for temozolomide (TMZ) resistance in GBM patients, though some conflicting data exist. We aimed to analyze methylation of the MGMT promoter and CHI3L1/CHI3L1/28 expression in cells of the panel of U-251 MG (U251) glioblastoma cell lines, sensitive and resistant to TMZ. Methods. Transgenic cell lines U251-GFP, U251-CHI3L1 and U251-CHI3L2 have been generated previously by lentiviral transduction. TMZresistant derivates of transgenic and parental U251 (tmzU251, tmzU251-GFP, tmzU251-CHI3L1 and tmzU251-CHI3L2) have been obtained by cells selection with augmenting concentrations of TMZ. Methyl-specific (MS) PCR and MS-sequencing were used for MGMT promoter methylation determination. qPCR with isoformspecific primers was carried out for the analysis of CHI3L1/ CHI3L1/18 expression. Results. Both MS-PCR and MSsequencing demonstrated an increase in MGMT promoter methylation in all the transgenic and the TMZ-resistant cell lines comparing to intact U251. MS-sequencing showed the most significant increase of methylation level in the tmzU251 cell line comparing to the original U251 (p < 0.001 by Kolmogorov-Smirnov test). TMZ also leads to an increase in methylation level of the MGMT promoter region in all the transgenic cell lines compared to the original U251 (p < 0.01). Transgenic cell lines except the U251-CHI3L1 demonstrated reduced expression of both CHI3L1 isoforms. On the contrary, TMZ long-term treatment leads to an increase of both CHI3L1 isoforms expression. Surprisingly the most significant rising was revealed in tmzU251-CHI3L2 with the isoforms ratio retention contrary to tmzU251 and tmzU251-GFP cells. Both these sublines showed bias towards CHI3L1 full isoform expression whereas tmzU251-CHI3L1 possessed balance shifting in favor of short CHI3L1A8 isoform. Conclusions. The obtained data indicate involvement of cellular mechanisms other than the reparative activity of MGMT in the evolution and maintenance of the TMZ-resistant phenotype. This is consistent with other studies on the decreasing of the prognostic role of the MGMT promoter methylation status for the assessment of therapeutic effect of TMZ in the treatment of glioblastoma. Balance alteration between the two CHI3L1 transcripts could be a part of the glioblastoma chemoresistance mechanism in response to TMZ treatment. Grants/Fundings: The work is supported by the Simons Support Grant 1290589.

**Keywords:** glioblastoma, temozolomide resistance, CHI3L1, MGMT.