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# Alternatively spliced short and long isoforms of adaptor protein intersectin 1 form complexes in mammalian cells

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*Intersectin 1 (ITSN1) is an adaptor protein involved in membrane trafficking and cell signaling. Long and short isoforms of ITSN1 (ITSN1-L and ITSN1-S) are produced by alternative splicing. The aim of our study was to investigate whether ITSN1-L and ITSN1-S could interact in mammalian cells. Methods. During this study we employed immunoprecipitation and confocal microscopy. Results. We have shown that endogenous ITSN1-S coprecipitates with overexpressed ITSN1-L in PC12, 293 and 293T cells. Long and short isoforms of ITSN1 also colocalize in 293T cells. Conclusions. ITSN1-L and ITSN1-S form complexes in mammalian cells.*

*Keywords: ITSN1, alternatively spliced isoforms, adaptor/scaffold proteins.*

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**Introduction.** ITSN1 is a multidomain and multifunctional adaptor protein which is involved in clathrin- and caveolin-dependent endocytosis [1, 2], Ca<sup>2+</sup>-regulated exocytosis [3] and synaptic vesicles retrieval [4]. It is also implicated in cellular signaling [5-7] and neuron survival [8]. Abnormalities of ITSN1 expression are associated with the endocytic anomalies reported in Down syndrome brains and early stages of Alzheimer's disease as well as with neurodegeneration in Huntington's disease [9, 10]. ITSN1 is also associated with glioma and neuroblastoma tumorigenesis [11, 12].

Two major ITSN1 isoforms are produced by alternative splicing [13]. Ubiquitously expressed ITSN1-S consists of two N-terminal EH (Eps15 homology) domains, coiled-coil region (CCR) and five SH3 (Src homology) domains. EH domains interact with NPF motifs and are highly involved in clathrin coated pits assembling [14], and SH3 domains interact with proline-rich PXXP motifs and provide protein-protein interactions

in many cellular processes, including membrane trafficking and signaling [15]. The long isoform of ITSN1 (ITSN1-L) is expressed predominantly in neurons and has three additional C-terminal domains: DH (dbl homology), PH (pleckstrin homology) and C2. The tandem of DH-PH domains is a GEF (guanine nucleotide exchange factor) for the Rho-family small GTPase Cdc42 [16].

ITSN1 domain composition implies that this is an adaptor/scaffold protein. These proteins possess multiple modular interaction domains and play a crucial role in spatial and temporal organization of cellular processes. Scaffolds regulate selectivity in signaling pathways using tethering mechanism and physically assembling chosen components of signaling pathway or network [17]. It is known that some scaffold proteins dimerize or even oligomerize to perform their function [18, 19]. As we have recently shown that ITSN1-S forms a heterodimer with its minor isoform 22a [20, 21], we presumed that probably other isoforms of ITSN1 are able to interact with each other thereby assembling protein supercomplexes.

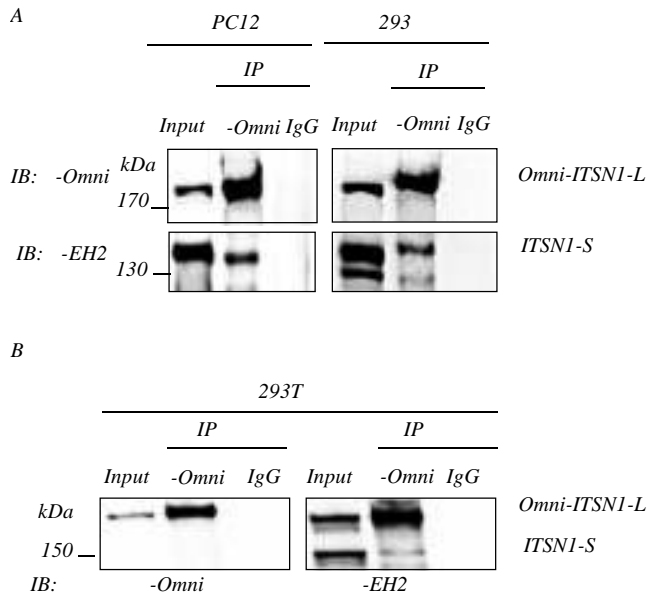


Fig. 1. ITSN1-L and ITSN1-S co-precipitate in mammalian cells. PC12 and 293 cells (A) or 293T cells (B) were transfected with Omni-ITSN1-L and subjected to immunoprecipitation (IP) with anti-Omni antibody or control IgG followed by immunoblotting (IB) with anti-EH2 (anti-ITSN1) antibody

**Materials and methods. Antibodies.** A monoclonal anti-Omni (D-8) antibody was from «Santa Cruz Biotechnology» (USA). Rabbit polyclonal antibodies against the EH2 domain of ITSN1 were described previously [22]. HRP-conjugated goat anti-mouse and anti-rabbit antibodies were purchased from «Invitrogen» (USA).

**DNA plasmid constructs.** The coding sequence of ITSN1-L was amplified from human embryonic brain cDNA and cloned into the *pcDNA4His/Max C* vector («Invitrogen»).

Plasmids encoding GFP-ITSN1-L and mCherry-ITSN1-S were described previously [23, 24].

**Cell culture and transfection.** 293 and 293T cells were obtained from the American Type Culture Collection and maintained in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10 % fetal calf serum («Sigma», USA), 50 U/ml penicillin and 100 mg/ml streptomycin. The cells were transiently transfected using JetPEI transfection reagent (Polyplus Transfection) according to manufacturer recommendations and further processed 24 h after transfection.

PC12 cells were maintained in DMEM supplemented with 5 % fetal calf serum («Sigma»), 10 % horse serum («Sigma»), 50 U/ml penicillin and 100 mg/ml streptomycin. The cells were transiently transfected using

Lipofectamine 2000 transfection reagent («Invitrogen») according to manufacturer recommendations and further processed 48 h after transfection.

**Immunoprecipitation and Western blot analysis.** The immunoprecipitation and Western blot were performed as described previously [25]. In brief, the cells were lysed in IP buffer (150 mM NaCl, 20 mM Tris, pH 7.5, 10 % glycerol, 0.5 % NP40, protease inhibitors cocktail («Sigma»)) and centrifuged for 15 min at 16,000 g. Supernatant (2 mg of proteins) was incubated with 2 µg anti-Omni antibodies and 20 µl Protein A/G Plus Ultralink Resin («Thermo Scientific», USA) for 4 h at 4 °C. Then beads were washed four times with IP buffer without inhibitors. Immunoprecipitated complexes were eluted with Laemmli buffer, resolved by SDS-PAGE and transferred to nitrocellulose membranes («Bio-Rad», USA). The membranes were blocked with 5 % non-fat milk in TBS-T (1 × TBS («Euromedex», France), 0,1 % Tween 20) for 1 h, incubated with anti-Omni or anti-EH2 antibodies and washed. Detection was performed by horseradish peroxidase-conjugated goat anti-mouse or anti-rabbit antibodies.

**Confocal microscopy.** 293T cells were transfected with fluorescent protein constructs, fixed in 4 % paraformaldehyde in PBS 24 h after transfection, washed two times with PBS and mounted in Mowiol medium («Sigma»). The slides were analyzed using Leica SP5 confocal microscope.

**Results and discussion.** In order to prove the existence of ITSN1 macromolecular complexes in cells, we have performed a co-immunoprecipitation assay in different mammalian cell lines (Fig. 1).

Omni-tagged ITSN1-L was overexpressed in rat pheochromocytoma PC12 cell line and in human 293 or 293T cell lines. The immunoprecipitation was carried out with anti-Omni antibodies and further immunoblotting was performed with anti-EH2 antibodies to detect precipitated ITSN1-S. As a control of non-specific binding appropriate amount of mouse IgG was used. In all tested cell lines endogenous ITSN1-S readily co-precipitated with recombinant ITSN1-L. 293T cell line was of particular interest as it expressed endogenous ITSN1-L at relatively high level in contrast to 293 cell line (data not shown). And in addition to full-size ITSN1-L protein in 293T cells a minor band with slightly lower molecular weight is present which is presumably

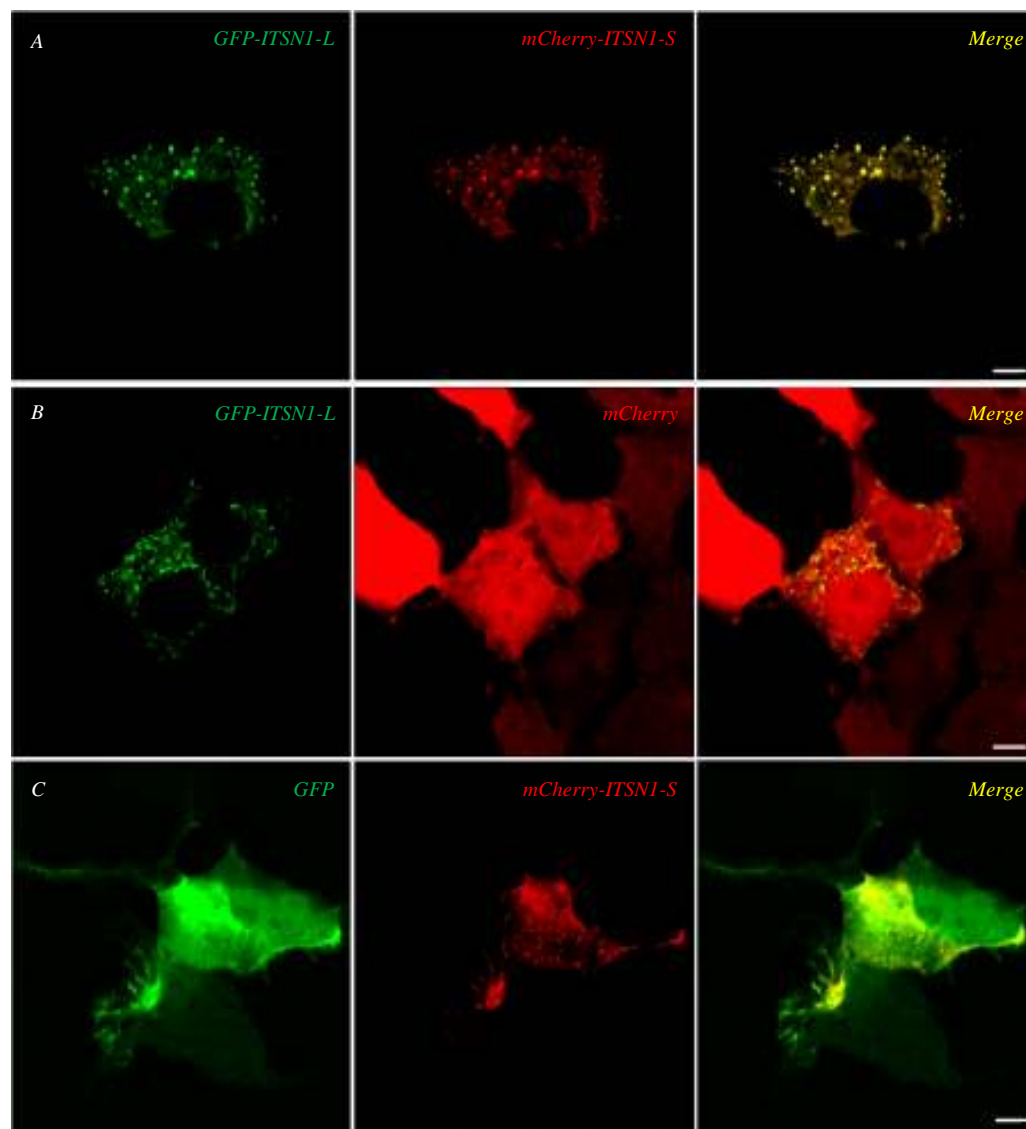


Fig. 2. Overexpressed GFP-ITSN1-L and mCherry-ITSN1-S colocalize in 293T cells. 293T cells were transfected with both GFP-ITSN1-L and mCherry-ITSN1-S (A) or with GFP-ITSN1-L alone and empty pmCherry vector (B) or with mCherry-ITSN1-S alone and empty pGFP-C1 vector (C). Scale bar represents 5  $\mu$ m

ly one of the possible alternatively spliced isoforms of ITSN1-L. The bands below the ITSN1-S can also refer to one of its minor isoforms, e. g. ITSN1-22a [20, 21]. And these minor isoforms also co-precipitate with overexpressed ITSN1-L suggesting that ITSN1 can form heterodimers of different isoform composition (Fig. 1, B). To explore whether ITSN1-S and ITSN1-L have similar subcellular localization, we overexpressed GFP-tagged ITSN1-L and mCherry-tagged ITSN1-S in 293T cells (Fig. 2, A).

As a control we co-expressed each of the proteins with empty vector (Fig. 2, B, C). Obtained data clearly indicate that both isoforms perfectly colocalize and have similar subcellular distribution.

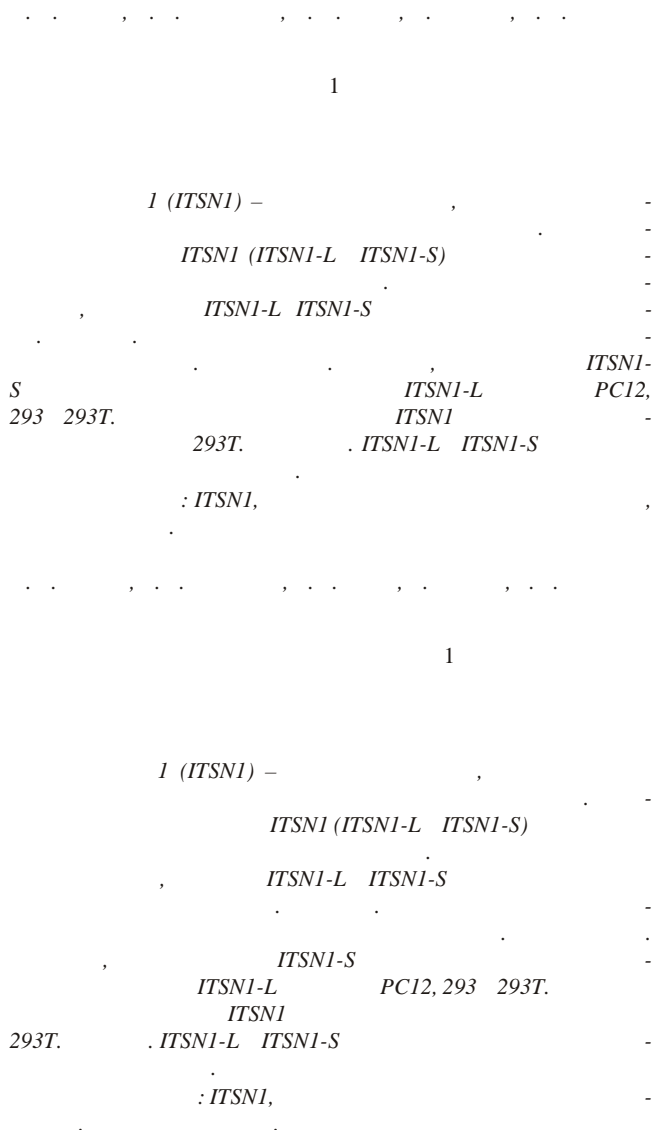
So we have demonstrated that ITSN1 isoforms interact in mammalian cells, thereby forming large complexes with varying protein composition that can function in membrane trafficking as well as in cell signaling. However the molecular mechanism controlling this clusterization remains unclear.

Such ITSN1-ITSN1 interaction possibly could be mediated by CCR as it is the case for Eps15, another CCR and EH domain containing protein [26]. This supposition is also supported by the work of Wong et al. [27]. They have used a high throughput yeast two hybrid screening to define the possible partners of the ITSN scaffolds and have identified ITSN1 as a target of a prey containing its EH2 domain and a half of CCR

(204–522 aa). They have further shown the existence of ITSNI homo- and heteromeric complexes by bimolecular fluorescence complementation.

But we assume that ITSNI SH3 domains may also contribute to this interaction as it has already been shown that some other proteins dimerize via their SH3 domains [28, 29]. These assumptions will be tested in our further research.

**Conclusions.** We have shown that ITSNI long and short isoforms form complexes in different mammalian cell lines.



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